

Region 10, 1200 Sixth Avenue, Seattle WA 98101

# COLUMBIA RIVER BASIN FISH CONTAMINANT SURVEY

# VOLUME I Appendix A

STUDY DESIGN FOR ASSESSMENT OF CHEMICAL CONTAMINANTS IN FISH CONSUMED BY FOUR NATIVE AMERICAN TRIBES IN THE COLUMBIA RIVER BASIN

January 2002

Prepared by
U.S. Environmental Protection Agency
Region 10
Seattle, Washington

Final	Draft	Re	port

Assessment of Chemical Contaminants in Fish Consumed by Four Native American Tribes in the Columbia River Basin

# Prepared For:

U.S. ENVIRONMENTAL PROTECTION AGENCY OFFICE OF WATER OFFICE OF SCIENCE AND TECHNOLOGY WASHINGTON, D.C.

# TABLE OF CONTENTS

FIGURES	ii
TABLES	iii
1.0 INTRODUCTION	<u>1</u>
1.1 PROJECT DESCRIPTION	<u>1</u>
1.2 DOCUMENT PURPOSE AND SCOPE	<u>1</u>
1.3 STUDY OBJECTIVES	<u>3</u>
2.0 STUDY DESIGN	<u>5</u>
2.1 SELECTION OF SAMPLING LOCATIONS	·
2.2 SELECTION OF SPECIES	<u>11</u>
2.3 SAMPLE TYPE	<u>11</u>
2.4 SAMPLING STRATEGY	<u>15</u>
2.5 TARGET ANALYTES	<u>19</u>
3.0 RECOMMENDATIONS	<u>28</u>
3.1 GENERAL COMMENTS	<u>28</u>
3.2 COMMENTS SPECIFIC TO THE PHASE II STUDY	<u>28</u>
40 REFERENCES	30

# **FIGURES**

	<u>Page</u>
Figure 1. Decision tree for selection of tissue sampling sites	6
Figure 2. Tribal fishing sites for resident fish where use exceeded forty percent	7
Figure 3. Tribal fishing sites for anadromous fish where use exceeded forty percent	8
Figure 4. Fish sampling locations	10
Figure 5. Phase II study design	13

# **TABLES**

			<u>Page</u>
Table 1.	Members of the task force for the CRITFC project		2
Table 2.	Percentage of adult tribal members consuming proposed target species and fishing sites where these species will be collected		12
Table 3.	Columbia River Inter-Tribal Fish Commission Exposure Study.  Adult consumption of fish parts		14
Table 4.	Study design for assessment of chemical contaminants in fish consumed by CRITFC member tribes		17
Table 5.	Columbia River Inter-Tribal Fish Commission Exposure Study. Study design for the Columbia River Inter-Tribal Fish Commission exposure study		18
Table 6.	Exposure assumptions for screening fish tissue chemical concentrations	20	
Table 7.	Chemicals that exceeded tissue screening concentrations		21
Table 8.	Chemicals that did not exceed tissue screening concentrations		26

## 1.0 INTRODUCTION

## 1.1 PROJECT DESCRIPTION

The Columbia River Inter-Tribal Fish commission (CRITFC) entered into a cooperative agreement with the U.S. environmental Protection Agency (EPA) in 1990 to conduct a fish consumption survey of the Nez Perce, Warm Springs, Umatilla, and Yakima Native American Tribes. This consumption survey, which was released in October 1994 (CRITFC 1994), was the first phase of a broader effort to determine the role of fish consumption as an exposure route for waterborne toxics among individuals of these tribes. The second phase will use the information from the consumption survey to design, and implement, a sampling program to collect tissue contaminant data from residence and anadromous fish species consumed by tribal members. It is this phase of the project with which this document is concerned. The third phase, which will determine blood contaminant levels of tribal members, has not been initiated. Collectively, these three components should provide the necessary information for developing an exposure assessment for members of the four CRITFC tribes. Information derived from this exposure assessment may then be used by U.S. EPA and others for developing an assessment of health risks to fish consumers in the four member tribes.

#### 1.2 DOCUMENT PURPOSE AND SCOPE

This scoping document was originally submitted in draft form to members of the CRITFC Task Force (Table 1), other tribal representatives, and selected government agencies. The draft document provided a preliminary study design that served as a starting point for discussions that occurred at a Design Conference held on October 19-20, 1994 in Portland, Oregon, at which a study design for U.S. EPA's Phase II CRITFC exposure study was finalized. This document presents the consensus study design developed at the Design Conference and provides the study objectives, rationale, and study recommendations formulated by attendees of the Design Conference. This document is not intended to be a sampling plan or a quality assurance/quality control plan. Such documents will be prepared prior to initiating the field sampling.

TABLE 1. MEMBERS OF	F THE TASK FORCE FOR THE CRITFC PROJECT
Name	Affiliation
Jeffrey Bigler, Project Manager	U.S. EPA Headquarters, Office of Water
Rick Albright	U.S. EPA Region X, Water Division
Harriett Amann	Washington Department of Health
Steve Bradbury	U.S. EPA Headquarters, Office of Research & Development
Pat Cirone	U.S. EPA Region X, Environmental Services Division
Dave Cleverly	U.S. EPA Headquarters, Office of Research & Development
Dana Davoli	U.S. EPA Region X, Environmental Service Division
Jerry Filbin	U.S. EPA Headquarters, Office of Policy, Planning and Evaluation
Gene Foster	Oregon Department of Environmental Quality
John Gabrielson	U.S. EPA Region X, Water Division
Clarice Gaylord	U.S. EPA Headquarters, Office of Environmental Equity
Jim Griggs	Warm Springs Tribe
Lynn Hatcher	Yakima Tribe
Gary James	Umatilla Tribe
Ken Kauffman	Oregon Department of Health
Craig McCormick	Washington Department of Ecology
Bruce Mintz	U.S. EPA Headquarters, Office of Water
Cynthia Nolt	U.S. EPA Headquarters, Office of Water
Brian Offord	Washington Department of Ecology
Carol Schuler	U.S. Fish and Wildlife Service
Anne Watanabe	10Columbia River Inter-Tribal Fish Commission
Silas Whitman	Nez Perce Tribe
Don Yon	Oregon Department of Environmental Quality

### 1.3 STUDY OBJECTIVES

The primary objectives of the Phase II study are to:

- Measure fish contaminant levels for species and fishing locations being utilized by CRITFC member tribes to provide, in conjunction with the CRITFC (1994) fish consumption report, an assessment of fish consumption as an exposure route for waterborne toxics among individuals of these tribes.
- To use the information derived from the exposure assessment to estimate potential health risks to fish consumers in the four CRITFC member tribes.

The objectives for the Phase II study were thoroughly discussed at the Design Conference and consensus was reached for the two primary objectives listed above. Specific details regarding how the collected data will be used to accomplish these objectives will be developed as the Phase II study progresses. Design Conference attendees recommended that the methodology for conducting an assessment of human health for the CRITFC member tribes be clearly delineated, as well as the form in which this information would be conveyed to the public. In particular, questions were raised about whether the data would allow only site-specific exposure assessment, or whether the data could be extrapolated to estimate exposure over larger areas of the Columbia River Basin. It was decided that this issue could not be fully resolved until it was determined whether contaminant levels varied significantly among different collection sites.

The manner in which human health concerns resulting from the Phase II study would be disseminated to the public was also discussed at the Design Conference. Concerns were raised by conference attendees about the potential differences in methodology and presentation of human health information by State Health departments, EPA, and other state regulatory agencies. Design Conference attendees recognized that different agencies would likely utilize the available data to meet their own specific operational mandates, and that the analyses and form of presentation of the data might differ. However, it was generally agreed that all agencies should

strive to keep each other informed about the uses and presentation of any data generated from the Phase II study.

Originally, a secondary objective of the Phase II study was to collect sediment contaminant data from the fish collection sites to aid in the determination of chemical-specific biota-sediment accumulation factors (BSAFs). While Design Conference attendees recognized the utility and merit of collecting this data, it was felt that available resources were insufficient to accomplish the primary objectives and carry out a statistically valid sampling program to determine BSAFs. Therefore, this secondary objective was eliminated in favor of a recommendation that additional resources be allocated, if possible, to determine BSAFs for the Columbia River Basin. Furthermore, there was general acknowledgment that implementation of this recommendation should be preceded by the development of a well planned, statistically valid, study design.

#### 2.0 STUDY DESIGN

This section provides a description and rationale for the study design developed for U.S. EPA's Phase II Columbia River Inter-Tribal Fish Commission (CRITFC) exposure study. The study design was developed through a consensus process that considered the objectives presented in Section 1.3. The information used in developing this study design included the fish consumption data provided in the CRITFC (1994), existing data on chemical concentrations in fish tissue within the Columbia River Basin collected from 1984 - 1994, and the results of a human health risk-based screening analysis of the existing data. The main constraint on the study design was the resources available for dioxin and PCB congener analysis of tissue and sediment samples (\$250,000).

## 2.1 SELECTION OF SAMPLING LOCATIONS

Figure 1 illustrates the decision process that was used to select the sampling sites for both resident and anadromous fish species. Initially, fishing sites that represented greater than 40 percent of each tribe's fishing use for resident and anadromous fish species were identified. The 22 fishing locations for resident species that met this criterion were located in the Clearwater, Deschutes, and Umatilla watersheds, and the mainstem Columbia River below McNary Dam (Figure 2). For anadromous species, the same 22 locations plus 4 additional sites located in the mainstem Columbia River upstream from the mouth of the Snake River to Rocky Reach Dam represented greater that 40 percent of the fishing use (Figure 3). To reduce the number of sites to a number consistent with the resources available for the Phase II sampling effort, the distribution of fishing sites exceeding the 40 percent use criterion was subdivided into two categories: watersheds with multiple fishing sites (i.e., Clearwater, Deschutes, and Umatilla), and mainstem Columbia River segments represented by a single fishing site (fishing sites 5-9, 15, 16, and 18). For the three watersheds with multiple fishing sites, a single site located near the base of the watershed (i.e., a second order river segment) was selected to be representative of other fishing sites within the watershed. The three sites that meet this criterion are fishing sites 98, 30, and

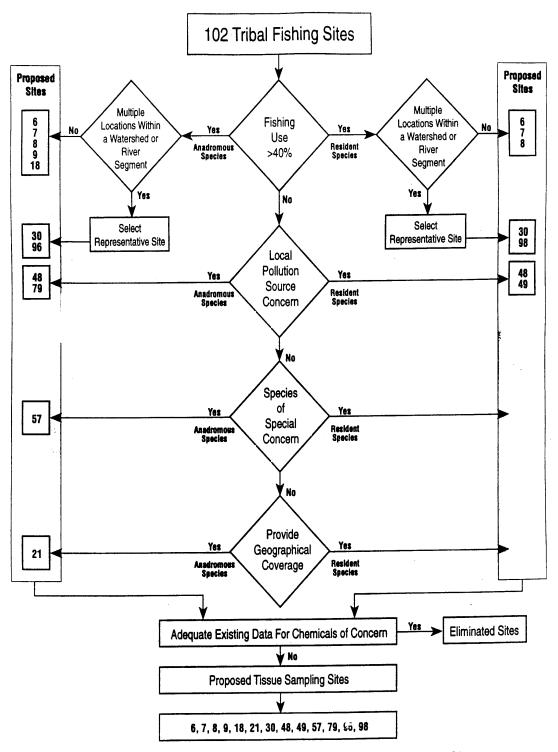
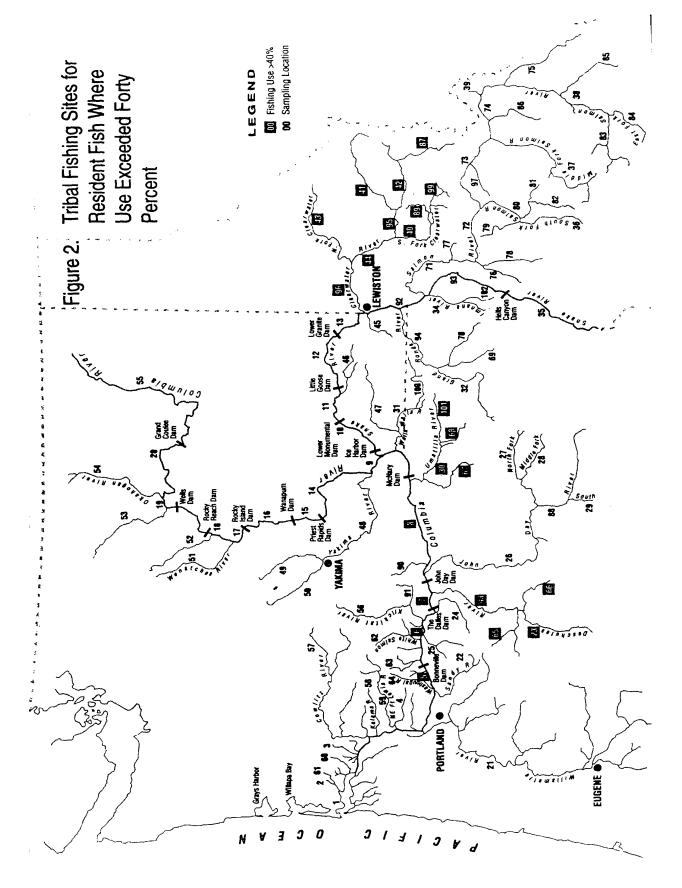
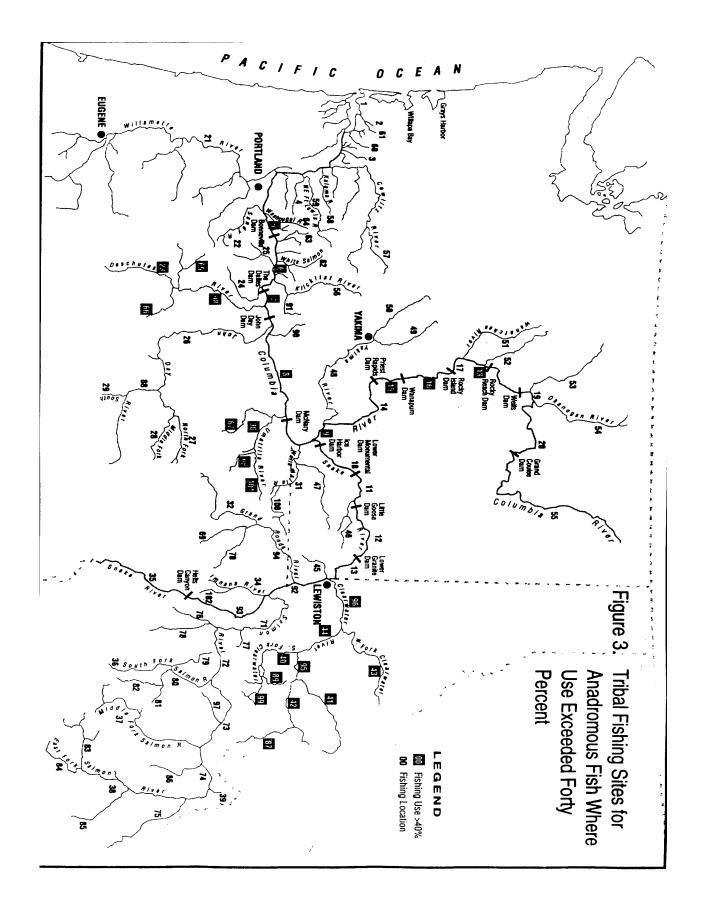


Figure 1. Decision Tree For Selection of Tissue Sampling Sites

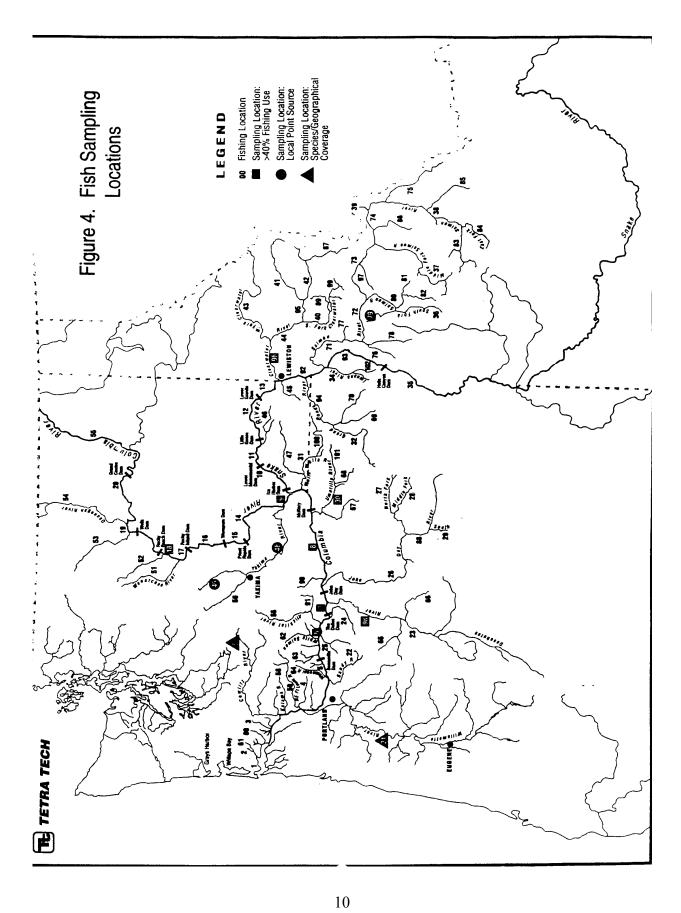




96 located in the Deschutes, Umatilla, and Clearwater Rivers, respectively (Figure 4). The assumption that these three sites will be representative of other fishing sites within each watershed is probably reasonable for anadromous species, but may not hold for resident species depending on local sources of contaminants and the ranges of the resident species being considered. The decision to analyze contaminant levels in resident species at the same sites as anadromous species was based on considerations of sampling logistics, and the desire to compare contaminant levels between both categories of fish.

Eight fishing sites in the mainstem of the Columbia River are located in river segments separated by dams. Sites 6, 7, and 8 were selected because they represented greater than 40 percent of the Yakima fishing use for both resident and anadromous species (Figure 4). Site 5, which also met these use criteria, was not selected because of the need to reduce the number of sampling locations, and because of the large amount of recent fish contaminant data that have been collected by Lower Columbia River Water Quality Bi-State Program in the vicinity of this site (Tetra Tech 1993; 1994a, b). Sites 9, 15, 16, and 18 are Columbia River mainstem sites that represent greater than 40 percent of the Yakima fishing use for anadromous species. Site 9 was selected by the Yakima representative due to its frequent use for fishing. Site 18 was selected because it represented the most upstream location with frequent fishing use. Fish collected at this site would presumably have the maximum exposure duration to contaminants within the mainstem Columbia River. Sites 15 and 16, which are located in the stretch of water between sites 9 and 18, were not selected because of the need to reduce the number of sites sampled.

Three sites (48, 49, and 79) were selected by tribal representatives because of concern about local pollution sources (Figure 4). Fishing use of these sites by tribal members is less than 20 percent. Sites 48 (Marion Drain) and 49 (Wilson Creek) are located in the Yakima River. There is concern that both of these sites have been adversely impacted from pesticide runoff (Hatcher, L., 28 September 1994, personal communication). Site 49 is also an important spawning site for rainbow trout. Site 79 is located in the Salmon River watershed in the vicinity of a mining operation.



The two remaining sites that are proposed for sampling were selected by considering a particular species of concern and the desire to provide broad geographical coverage of sampling sites (Figure 4). Site 57, in the Cowlitz River, was selected to provide contaminant data for smelt. Fifty-two percent of adult tribal members consume smelt (CRITFC 1994). Because this fish species has a high oil content, it may accumulate higher levels of hydrophobic organic contaminants than other anadromous species; therefore, CRITFC Task Force members felt it was important to include sampling at site 57. Site 21, in the Willamette River, was selected to provide additional geographic coverage, and to provide contaminant data for lamprey, which are consumed by 54 percent of adult tribal members (CRITFC 1994).

# 2.2 SELECTION OF SPECIES

The selection of species to be analyzed was based primarily on consumption data presented in CRITFC (1994). Table 2 shows the fish species that are consumed by tribal members and the proposed fishing sites where the species will be collected. Tissue samples for all consumed species except squawfish and shad will be analyzed. These two species are consumed by only a small fraction (<2.7 percent) of adult tribal members.

## 2.3 SAMPLE TYPE

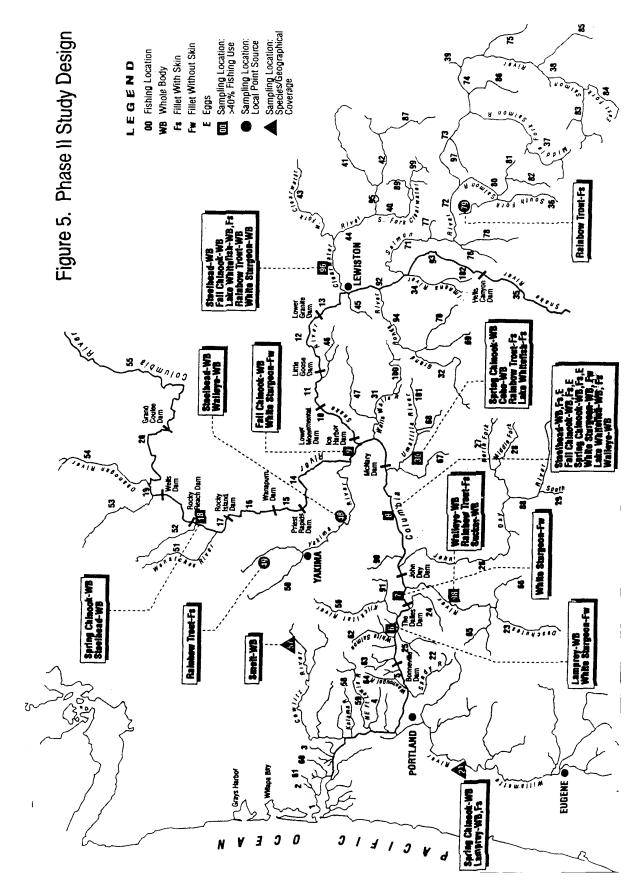
Figure 5 shows the locations, species, and sample types that will be analyzed during EPA's Phase II study. Four types of samples will be analyzed: whole-body (WB), fillet with skin ( $F_s$ ), fillet without skin ( $F_w$ ), and eggs (E). Whole-body samples were selected for several species to maximize the chances of measuring detectable levels of contaminants of concern and because data presented in CRITFC (1984) show that tribal members may consume several fish parts in addition to the fillet (Table 3). Eggs from spring chinook, fall chinook, and steelhead will be analyzed because consumption data shows that salmonid eggs are widely consumed by tribal members (Table 3). Because of the high lipid levels in eggs, concentrations of hydrophobic organic chemicals may reach substantially higher levels than in other fish tissues. Design Conference attendees felt that it was important to determine contaminant levels in various fish

TABLE 2. PERCENTAGE OF ADULT TRIBAL MEMBERS CONSUMING PROPOSED TARGET SPECIES AND FISHING SITES WHERE THESE SPECIES WILL BE COLLECTED

Species	Weighted Percent That	Proposed Fi	shing Sites
	Consume the Species	Site Numbers	Site Locations (Rivers)
Salmon	92.4%	21, 8, 9, 18, 30, 96	Willamette, Columbia, Umatilla, Clearwater
Lamprey	54.2%	21, 6	Willamette, Columbia
Trout <sup>a</sup>	70.2%	98, 8, 18, 30, 48, 49, 96,79	Deschutes, Columbia, Umatilla, Yakima, Clearwater, Salmon
Smelt	52.1%	57	Cowlitz
Whitefish	22.8%	8, 30, 96	Columbia, Umatilla, Clearwater
Sturgeon	24.8%	6, 7, 8, 9, 96	Columbia, Clearwater
Walleye	9.3%	98, 8, 48	Deschutes, Columbia, Yakima
Sucker	7.7%	98	Deschutes
Squawfish	2.7%	none	none
Shad	2.6%	none	none

Source: Modified from CRITFC (1994).

<sup>a</sup> Rainbow Trout and Steelhead.



IABLE 3. CULUMBIA RIVER INTER-TRIBAL FISH COMMISSION EXPOSURE STUDY. ADULT CONSUMPTION OF FISH PARTS		Bones Organs	Weighted % Weighted % That Consume	12.1% 470 3.7%	5.2% 250 3.2%	7.1% 362 2.3%	28.4% 206 27.9%	6.0% 124 0.0%	2.6% 121 0.3%	2.4% 46 0.9%	5.9% 15 0.0%	9.8% 42 2.1%	3.3% 15 0.0%	
RE STUDY. ADUI		Eggs	Weighted % That Consume	3 42.8%	0 4.6%	4 8.7%	9 46.4%	5 20.6%	1 11.9%	9.8%	11.1%	30.4%	5 0.0%	
XPOSUI	Parts		Z ge	473	250	38	200	125	121	99	51	42	92	
OMMISSION E		Head	Weighted % That Consume	42.7%	18.1%	13.7%	37.4%	15.4%	6.2%	6.2%	8.1%	19.4%	0.0%	
FISH CO			z	473	250	365	210	125	121	46	15	42	91	
INTER-TRIBAL		Skin	Weighted % That Consume	55.8%	89.3%	68.5%	88.9%	53.8%	18.2%	20.7%	34.1%	\$0.0%	15.7%	
A RIVER			Z	473	251	365	500	124	121	94	15	42	91	
LE 3. CULUMBI		Fillet	Weighted % That Consume	95.1%	86.4%	89.4%	78.8%	93.8%	%9.7%	100%	89.7%	89.3%	93.5%	<b>t</b> ).
I AB			Z	473	249	365	209	125	121	46	15	42	16	TFC (1994
			Species	Salmon	Lamprey	Trout	Smelt	Whitefish	Sturgeon	Wallcye	Sucker	Squawfish	Shad	Source: CRITFC (1994)

parts (i.e., whole-body, fillet, and eggs) so that this information could be used to provide guidance on how to prepare fish, or what parts should be avoided, in the event that contaminant levels exceed levels that warrant concern. In addition, the conversion factors developed from this data (e.g., whole-body: fillet and whole-body; egg ratios) may assist in the comparison of Phase II data with other historical data that exist within the Columba River Basin. Figure 5 indicates that most of the comparisons of contaminant levels in different fish samples will occur at Site 8 in the Columbia River between the McNary and John Day dams. This site was selected because of its importance as a fishing site for all four CRITFC member tribes.

# 2.4 SAMPLING STRATEGY

The sampling strategy proposed for this study design is consistent with guidance provided in the document entitled: *Guidance for Assessing Chemical Contaminant Data for Use in Fish Advisories*, Volume I: Fish Sampling and analysis (U.S. EPA 1993b). For all fish species except white sturgeon, three replicate composite samples will be analyzed from each collection site. For white sturgeon, one sample from three individual fish will be analyzed from each collection site. The number of fish per composite will likely vary for different species: 20 individuals per composite for smelt and lamprey, 8 individuals per composite for resident species, and 5 individuals per composite for salmon and steelhead (Table 4). U.S. EPA (1993b) recommends that 3 to 10 individuals should be collected for a composite sample for each target species and that the same number of individual organisms should be used to prepare all replicate composite samples for a given target species at a given site. Several ongoing fish contaminant studies in the Columbia river Basin are compositing 8 individuals per sample, so the use of this number would simplify comparisons with other available data. Because of the small size of lamprey and smelt, a composite of 8 individuals would not provide enough tissue for all chemical analyses; therefore a nominal value of 20 individuals per composite was suggested for these species. Design

Conference attendees felt that the number of individuals per composite for salmon and steelhead should be reduced from 8 to 5 (some individuals suggested 3) because of concerns about the ability to collect sufficient numbers of fish, and because it was felt that the study should strive to minimize impacts on these fish stocks.

Collection periods for each species have been tentatively assigned and are given in Table 5.

According to U.S. EPA guidance (U.S. EPA 1993b), the collection period should ideally avoid the spawning period of the target species, because many fish are subject to stress during spawning. However, because eggs will be collected from salmonid species, the typical spawning period for these species will be targeted (WDF/ODFW 1993). For resident species, wide collection periods have been proposed so that spawning periods can be avoided (Table 5). For white sturgeon, the proposed collection period is consistent with seasons established in previous years (WDF/ODFW 1994).

	TABLE 4. STUDY	DESIGN FC	OR ASSESSMENT OF C	HEMICAL CONTA	STUDY DESIGN FOR ASSESSMENT OF CHEMICAL CONTAMINANTS IN FISH CONSUMED BY CRITFC MEMBER TRIFFS	NSUMED BY CRITFC	MEMBER TRIBES	
Species	Classification	Fishing Site <sup>a</sup>	Waterbody	Sample Type <sup>b</sup>	Number of Composite Samples	Number of Individual Samples	Number of Fish per Composite	Total Number of Fish
Smelt	Anadromous	57	Cowlitz River	WB	3		20	09
Lamprey	Anadromous	21 6	Willamette River Columbia River	WB, Fs WB	3	1 1	20	120
Steelhead	Anadromous	8 18 8 9 9 9 9 9 9 9 9 9 9 9 9 9 9 9 9 9	Columbia River Columbia River Yakima River Clearwater River	WB, Fs, E WB WB WB	9 11 11 11	1 1 1 1	80 80 80 8	30 <sup>c</sup> 15 15
Coho	Anadromous	30	Umatilla River	WB	3	1	5	15
Chinook (Fall)	Anadromous	8 6 8	Columbia River Columbia River Clearwater River	WB, Fs, E WB WB	<b>0</b>	111	אטא	30 <sup>c</sup> 15
Chinook (Spring)	Anadromous	21 8 18 30	Willamette River Columbia River Columbia River Umatilla River	WB WB, Fs, E WB WB	m 04 m m	1111	<i>2</i> 0 <i>2</i> 0 <i>2</i> 0 <i>2</i> 0	15 30 <sup>c</sup> 15
Rainbow Trout	Resident	98 49 30 79	Deschutes River Yakima River Clearwater River Umatilla River Salmon River	Fs Fs WB, Fs Fs	ттчтт	1111	o ∞ ∞ ∞ ∞	2 4 4 4 8 4 2 6
White Stargeon	Resident	2 × × × × × × × × × × × × × × × × × × ×	Columbia River Columbia River Columbia River Columbia River Clearwater River	Fw Fv WB, Fw Fw WB	1111			33333
Lake Whitefish	Resident	8 98	Columbia River Umatilla River Clearwater River	WB, Fs Fs WB	<b>9</b> 6 6	111	00 00 00	24 24 24
Largescale Sucker	Resident	86	Deschutes River	WB	3		88	24
Wallcye	Resident	98 48 8	Deschutes River Yakima River Columbia River	WB WB	. E. E. E.	111	∞ ∞ ∞	24
TOTALS		13 sites	8 Rivers	4 Sample Types	105	18		618
A Nominal collection  B WB = Whole bod  C Assumes eggs will	1 areas. ly, Fs = Fillet with sk be removed from wh	cin, Fw = Fi	A Nominal collection areas.  b WB = Whole body, Fs = Fillet with skin, Fw = Fillet without skin, E = Eggs.  c Assumes eggs will be removed from whole-body samples and analyzed separately	ggs. tely.				

			TABLE 5. COL	UMBIA RIVER	INTER-TRIBAL	COLUMBIA RIVER INTER-TRIBAL FISH COMMISSION EXPOSURE STUDY	POSURE STUDY		
			Study Desi	gn for the Colu	mbia River InterTri	Study Design for the Columbia River InterTribal Fish Commission Exposure Study	posure Study		
Site	<u>u</u>	Receiving Water	Anadromous Fish Species (Sex)	Sample Type <sup>b</sup>	Collection Period <sup>a</sup>	Resident Fish Species (Sex)	Sample Type <sup>b</sup>	Collection Period	Number of Samples at Site <sup>C</sup>
48		Yakima Ríver	Steelhead (F)	WB	Dec-June	Walleye	WB	Jan-Dec	9
18		Columbia River	Spring Chinook (F) Steethead (F)	WB WB	Sept-Nov Dec-June	1	1	1	9
6		Columbia River	Fall Chinook (F)	WB	Sept-Oct	White Surgeon	Fw	Jan-Mar	9
57		Cowlitz River	Smelt (F)	WB	Feb	ı	ı	1	3
9		Columbia River	Lamprey	WB	June-July	White Sturgeon	Fw	Jan-Mar	9
7		Columbia River			1	White Stargeon	Fw	Jan-Mar	3
∞		Columbia River	Spring Chinook (F) Fall Chinook (F) Steelhead (F)	WB, Fs, E WB, Fs, E WB, Fs, E	Sept-Nov Sept-Oct Dec-June	White Sturgeon Lake Whitefish Walleye	WB, Fw WB, Fs WB	Jan-Mar Jan-Dec Mar-June	42
6 <del>4</del>		Yakima River		:		Rainbow Trout	Fs	Mar-June	3
8		Clearwater River	Steelhead (F) Fall Chinook (F)	WB WB	Dec-June Sept-Oct	Lake Whitefish Rainbow Trout White Sturgeon	WB, Fs WB WB	Late July Mar-June Jan-Mar	81
79		Salmon River	-	-	1	Rainbow Trout	Fs	Mar-June	3
93		Umatilla River	Spring Chinook (F) Coho (F)	WB WB	Sept-Nov Oct-Dec	Rainbow Trout (F) Lake Whitefish (F)	FS FS	June-Dec Jan-Dec	12
86		Deschutes River	-	-	<b>!</b>	Rainbow Trout Largescale Sucker Walleye	Fs WB WB	Mar-June Jan-Dec Mar-June	6
21		Willamette River	Spring Chinook (F) Lamprey	WB WB, Fs	Sept-Nov June-July	-	;		6
								Total + 12QA	126
MB.	= Who	WB = Whole body, Fs = Fillet with skin,		Fw = Fillet without skin, E = Eggs.	: = Eggs.				

WB = Whote body, Fs = Fillet win skin, FW = Fillet winkni skin, E = Eggs.

A Dates reflect typical spawning times. Source: Status Report. Columbia River Fish Runs and Fisheries 1938-1992 (WDF/ODFW 1993).

B Samples from all species except surgeon are composites from 5-20 individuals. Surgeon samples are from individual fish.

C Number of samples assumes each tissue sample performed in triplicate, each sediment sample performed in quadruplicate.

#### 2.5 TARGET ANALYTES

Target analytes were selected by considering the guidance provided in U.S. EPA (1993b) and by performing a health risk-based screening analysis of tissue contaminant data collected within the Columbia River Basin during the last ten years (1984-1994). The exposure assumptions used to perform the screening analysis are given in Table 6. Screening for carcinogens was performance for a 70 kg adult using a target cancer risk of 1 x 10<sup>-6</sup>. Screening for non-carcinogens was performed for a 14.5 kg child using a target hazard quotient of 0.1. Fish consumption rates assumed for adults and children were 194 and 81 g/day, respectively, which correspond to the cumulative 97<sup>th</sup> percentile consumption rate reported in CRITFC (1994). For chemicals that had both slope factors for estimating carcinogenic risk and reference does for estimating non-carcinogenic risk, separate tissue screening concentrations (STCs) were calculated and the lower of the two values was used for the screening analysis. Chemical concentrations reported as not detected were assumed to be equal to one half the detection limit for the screening analysis.

Table 7 lists the chemicals that exceeded tissue screening concentrations (STCs) and the frequency of exceedances. Chemicals that exceeded STCs include dioxins/furans, PCBs, organochlorine and organophosphorus pesticides, PAHs and other semivolatiles, trace metals, and radionuclides. Table 8 provides a list of the chemicals that did not exceed STCs. It should be noted that the tissue screening analysis could only be conducted for chemicals that have established slope factors or reference doses; therefore, Table 8 includes chemicals that do not have either of these toxicological reference values.

The final list of chemicals that will be analyzed during the Phase II study will be presented in a sampling and QA/QC plan that will be prepared prior to initiating sampling. This document will also provide the analytical methods and quantitation levels expected for the laboratory analyses. The chemical groups expected to be analyzed and a preliminary listing of the methods that may be

employed are provided below:

TABLE 6. EXPOSURE ASSUMPTION	NS FOR SCREENING FIS	SH TISSUE CHEMICAL
CON	CENTRATIONS	
	Cancer	Non-Cancer
Target Cancer Risk	1 x 10 <sup>-6</sup>	
Target hazard quotient		0.1
Body weight - Adult (kg)	70	
Body weight - Child (kg)		14.5
Averaging time - Adult (years of life)	70	
Averaging time - Child (years of life)		10
Exposure frequency (days/year)	365	365
Fish ingestion rate - Adult (grams/day)	194 <sup>b</sup>	
Fish ingestion rate - Child (grams/day)		81 <sup>b</sup>

Oral carcinogenic slope factors and oral reference doses were obtained from IRIS or HEAST.

<sup>&</sup>lt;sup>a</sup> This value is the 97<sup>th</sup> percentile consumption rate for fish consumers cited in Phase I of this project. CRITFC (1994). Table 10.

<sup>&</sup>lt;sup>b</sup> This value is the 97.4th percentile consumption rate for fish consumers cited in Phase II of this project. CRITFC (1994). Table 24.

	1		1	T	Τ	Т	T		T-	<del>-</del>	T	T -	Ī	T	T	T	T	_	T-	<del>-</del>	Γ	<u> </u>	T	T	T					
	Frequency	or SIC Exceedances	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	%0.001	100.0%	100.0%	89.66	94.6%	96.5%	94.8%	94.5%	76.6%	74.2%	54.2%	53.1%	4.4%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	100.0%	99.4%
	Total Number	or SIC Exceedances	264	10	265	265	265	229	228	548	541	228	188	221	182	172	36	135	104	121	8	109	220	220	187	33	143	231	268	326
IONS	Maximum	Concentration (μg/kg) <sup>a</sup>	0.09172	0.001385	0.01885	0.049	0.00336	600.0	0.0115	0.05602	0.32069	0.01902	0.0045	0.05432	0.0056	0.003	0.001	1	0.0055	0.002665	0.036	25	100	40	121	26	100	2700	1403	2043.1
NCENTRAT	Number	Detected	142	0	54	11	38	19	73	323	511	80	37	79	37	34	4	142	50	25	61	0	0	1	2	0	-	74	82	132
CREENING CC	Total	Measurements	264	10	265	265	265	229	228	548	541	229	192	229	192	182	47	182	192	228	182	109	220	220	187	33	143	231	897	328
EEDED TISSUE S (Page 1 of 5)	J.Lo	Classification <sup>b</sup>	ပ	٥	၁	၁	၁	၁	၁	C	၁	၁	၁	၁	၁	ى د	၁	၁	၁	၁	၁	၁	၁	၁	o o	С	C	С	C	ပ
LE 7. CHEMICALS THAT EXCEEDED TISSUE SCREENING CONCENTRATIONS (Page 1 of 5)	Screening Tissue	(μg/kg) <sup>a</sup>	0.000024	0.000024	0.000024	0.000024	0.000024	0.000005	0.000024	0.000002	0.000024	0.000005	0.000024	0.000048	0.000024	0.000024	0.000058	0.0024	0.00024	0.00024	0.0024	0.047	0.047	0.047	0.047	0.047	0.047	0.047	0.047	0.047
TABLE 7. CHEN		Chemical	1,2,3,4,6,7,8-HpCDD	1,2,3,4,6,7/1,2,3,4,7,8-HxCDF	1,2,3,4,7,8-HxCDD	1,2,3,6,7,8-HxCDD	1,2,3,7,8,9-HxCDD	1,2,3,7,8-PeCDD	2,3,4,6,7,8-HxCDF	2,3,7,8-TCDD	2,3,7,8-TCDF	2,3,4,7,8-PeCDF	1,2,3,7,8,9-HxCDF	1,2,3,7,8-PeCDF	1,2,3,6,7,8-HxCDF	1,2,3,4,7,8-HxCDF	TOTAL HxCDD	осрр	1,2,3,4,6,7,8-HpCDF	1,2,3,4,7,8,9-HpCDF	OCDF	Aroclor 1016	Aroclor 1221	Aroclor 1232	Aroclor 1242	Aroclor 1242/1016	Aroclor 1248	Aroclor 1254	Aroclor 1260	TOTAL PCBs
		Chemical Group	Dioxins/furans					けったっこ														PCBs								

	TABLE 7. CHE	TABLE 7. CHEMICALS THAT EXCEEDED TISSUE SCREENING CONCENTRATIONS  (Page 2 of 5)    Screening Tissue	EEDED TISSUE S (Page 2 of 5)	CREENING CO	NCENTRAT	TONS	Total Number	Frequency
Chemical Group	Chemical	Concentration (STC) (μg/kg) <sup>a</sup>	STC Classification <sup>b</sup>	Total Measurements	Number Detected	Concentration (µg/kg) <sup>a</sup>	of STC Exceedances	of STC Exceedances
Pesticides	Aldrin	0.021	၁	269	10	103	269	100.0%
Organocino inco	alpha-BHC	0.057	၁	433	28	39	433	100.0%
	beta-BHC	0.20	S	412	14	150	412	100.0%
	Chlordane	0.28	ပ	184	3	0/	184	100.0%
	Chlordane (tech)	0.28	၁	23	18	144	23	100.0%
	Dicofol	0.82	၁	186	17	300	186	100.0%
	Dieldrin	0.023	၁	478	113	352	478	100.0%
	gamma-BHC	0.28	၁	425	27	50	425	100.0%
	gamma-Chlordane	0.28	၁	249	40	09	249	100.0%
	Heptachlor	0.080	S	287	22	89	287	100.0%
	Heptachlor epoxide	0.040	၁	431	17	20	431	100.0%
	Lindane	0.28	၁	5	0		5	100.0%
	o,p'-DDE	1.1	၁	313	43	65	313	100.0%
	o,p'-DDT	1.1	၁	313	42	105	313	100.0%
	Pentachlorophenol	3.0	С	129	0	0009	129	100.0%
	TOTAL BHC	0.28	၁	29	3	160	29	100.0%
	Total Chlordane	0.28	၁	29	18	200	29	100.0%
	TOTAL DDT	1.1	၁	09	57	3000	99	100.0%
	Toxaphene	0.33	ى ن	311	38	1200	311	100.0%
	Hexachlorobenzene	0.23	C	334	24	250	333	99.7%
	alpha-Chlordane	0.28	၁	234	58	20	233	89.66
	Endosulfan II	0.90	N	227	7	90	214	94.3%
	Endosulfan I	06:0	Z	227	12	148	213	93.8%
	p,p'-DDE	1.1	С	498	391	3400	466	93.6%
	p,p'-DDT	1.1	C	473	215	096	424	%9.6%
	o,p'-DDD	1.5	C	313	72	130	262	83.7%
	DDD-'q,q	1.5	S	430	214	420	330	76.7%
	Endosulfan	3.6	z	49	9	170	36	73.5%

	TABLE 7. CHE	E 7. CHEMICALS THAT EXCEEDED TISSUE SCREENING CONCENTRATIONS (Page 3 of 5)	EEDED TISSUE S (Page 3 of 5)	CREENING CO	NCENTRAT	IONS		
		Screening Tissue				Maximum	Total Number	Frequency
Chemical Group	Chemical	Concentration (STC) (µg/kg) <sup>a</sup>	STC Classification <sup>b</sup>	Total Measurements	Number Detected	Concentration (µg/kg) <sup>a</sup>	of STC Exceedances	of STC Exceedances
Pesticides (Cont.)	Mirex	3.6	z	262	5	50	180	68.7%
	Endrin	5.4	z	467	17	19	59	12.6%
	Methoxychlor	89.5	z	240	10	832	6	3.8%
Organophosphate	Methyl parathion	4.5	z	106	9	38	55	51.9%
Semi-Volatiles	2,4,6-Trichlorophenol	32.8	၁	129	0	1250	129	100.0%
	2,4-Dichlorophenol	53.7	Z	129	0	750	129	100.0%
	2,4-Dinitrophenol	35.8	Z	128	0	2500	128	100.0%
	2,4-Dinitrotoluene	35.8	Z	129	1	1250	129	100.0%
	2,6-Dinitrotoluene	0.53	၁	129	0	1250	129	100.0%
	3,3'-Dichlorobenzidine	08.0	ວ	129	0	1250	129	100.0%
	Benzidine	0.0016	Э	23	0	815	23	100.0%
	Bis(2-chloroethy1)ether	0.33	o o	258	0	250	258	100.0%
	Bis(2-chloroisopropyl)ether	5.2	၁	258	0	250	258	100.0%
	Bis(2-ethylhexyl)phthalate	25.8	၁	129	51	34200	129	100.0%
	Hexachlorocyclopentadiene	125.3	Z	127	0	1305	127	100.0%
	Hexachloroethane	17.9	၁	129	0	250	129	100.0%
	N-Nitroso-di-n-propylamine	0.052	C	96	1	2900	96	100.0%
	N-Nitrosodimethylamine	0.0071	၁	23	0	139	23	100.0%
	Nitrobenzene	9.0	z	129	0	250	129	100.0%
	Pyridine	17.9	Z	23	0	139	23	100.0%
	Hexachlorobutadiene	4.6	၁	146	. 0	250	128	87.7%
	Carbazole	18.0	၁	73	0	250	99	76.7%
	4-Chloroaniline	71.6	Z	54	. 0	750	35	64.8%
	2-Chlorophenol	89.5	Z	129	1	4200	11	8.5%
	4-Nitrophenol	1109.9	z	129	2	4000	8	6.2%
	1,2,4-Trichlorobenzene	179.0	z	147	9	3100	6	6.1%
	Isophorone	379.8	၁	105	7	430	1	1.0%
	2,4-Dimethylphenol	358.0	z	129	0	909	1	0.8%
	Butyl benzyl phthalate	3580.2	z	129	1	3700	1	0.8%
	Di-n-octylphthalate	358.0	z	129	1	2640	1	0.8%

Chemical Concentration (STC)         STC         Total Concentration (STC)         STC         Total Concentration (STC)         Concentration (STC) <th< th=""><th></th><th>TABLE 7. CHE</th><th>TABLE 7. CHEMICALS THAT EXCEEDED TISSUE SCREENING CONCENTRATIONS (Page 4 of 5)</th><th>EEDED TISSUE S</th><th>CREENING CO</th><th>NCENTRAT</th><th>IONS</th><th></th><th></th></th<>		TABLE 7. CHE	TABLE 7. CHEMICALS THAT EXCEEDED TISSUE SCREENING CONCENTRATIONS (Page 4 of 5)	EEDED TISSUE S	CREENING CO	NCENTRAT	IONS		
PAHS   Benzi   Concentration (STC   Total   Number   Concentration (STC   Classification   Measurements   Concentration   Concentration   Classification   Classification   Concentration   Classification   Classification   Concentration   Classification   Clas			Screening Tissue				Maximum	Total Number	Frequency
PAHS         Benzolalparthracene         0.34         C         146         0         100           Benzolalpyrene         0.049         C         328         2         700           Benzolalpyrene         0.40         C         33         0         5           Benzolalfuoranthene         0.40         C         131         1         800           Chrysene         0.049         C         131         1         700           Chrysene         0.049         C         146         0         200           Chrysene         0.049         C         146         0         200           Divence         0.049         C         146         0         200           Berrollam         0.049         C         146         0         200           Berrollam         0.18         C         146         0         200           Accnaphthene         1074.1         N         147         3         3800           Accnaphthene         1074.1         N         147         3         3200           Accnaphthene         1074.1         N         145         45         2420           Marganese         89.	Chemical Group	Chemical	Concentration (STC)	STC Classification <sup>b</sup>	Total Measurements	Number Detected	Concentration (µg/kg) <sup>a</sup>	of STC Exceedances	of STC Exceedances
Benzo(a)pyrene         0.049         C         338         2         700           Benzo(b)fluoranthene         0.40         C         33         0         5           Benzo(b)fluoranthene         0.40         C         131         1         800           Chrysene         0.049         C         163         0         100           Chrysene         0.049         C         146         0         200           Dhenz[a,h]archene         0.049         C         146         0         200           Berto(lg,h)iperylene         2.3         C         146         0         200           Berto(lg,h)iperylene         2.3         C         146         0         200           Acenaphthene         1074.1         N         147         3         380           Acenaphthene         1074.1         N         147         3         380           Arenic         0.21         C         162         0         20           Arenic         0.21         C         162         0         20           Marganese         89.5         N         45         45         2420           Mariannese         5370	Semi-Volatile PAHs	Benz[a]anthracene	0.34	S	146	0	100	146	100.0%
Betrzo(b,k fh\u00erantheneeneeneeneeneeneeneeneeneeneeneeneene	<b>⇒</b>	Benzo[a]pyrene	0.049	၁	328	2	700	328	100.0%
Bernzollylihoranthene         0,40         C         131         1         800           Chrysene         0,93         C         131         1         700           Chrysene         0,049         C         163         0         100           Dibenzla, hlanthracene         0,045         C         146         0         200           Indenol1, 2,3-cdlpyrene         0,18         C         146         0         200           Acenaphthene         1074.1         N         147         3         3800           Acenaphthene         1074.1         N         147         3         3800           Arenic         0,21         C         162         0         200           Arenic         0,21         C         267         126         1860           Beryllium         0,084         C         40         2         60           Manganese         89.5         N         45         45         24200           Mercury         5.4         N         335         321         100000           Lead         7.7         N         296         13600           Selenium         90         N         207 <th></th> <td>Benzo[b,k]fluoranthene</td> <td>0.40</td> <td>U</td> <td>33</td> <td>0</td> <td>5</td> <td>33</td> <td>100.0%</td>		Benzo[b,k]fluoranthene	0.40	U	33	0	5	33	100.0%
Benzo(k)filuoranthene         0.949         C         131         1         700           Chrysene         0.049         C         163         0         100           Dibenz(a,h)anthracene         0.045         C         146         0         200           Indeno[1,2,3-cd]pyrene         0.18         C         146         0         200           Benzo[g,h,i]perylene         2.3         C         146         0         200           Acenaphthene         1074.1         N         147         3         3800           Arsenic         0.21         C         162         0         200           Arsenic         0.21         C         267         126         1860           Beryllium         0.084         C         40         2         60           Manganese         89.5         N         45         24200           Mercury         5.4         N         45         24200           Mercury         5.4         N         236         136000           Shenium         9.0         N         207         20         2200           Antimony         7.2         N         172         47200 <th></th> <td>Benzo[b]fluoranthene</td> <td>0.40</td> <td>U</td> <td>131</td> <td>-</td> <td>908</td> <td>131</td> <td>100.0%</td>		Benzo[b]fluoranthene	0.40	U	131	-	908	131	100.0%
Chrysene         0.049         C         163         0         100           Dibenz(a,h]anthracene         0.045         C         146         0         200           Indenol1.2.3-cdlpytrene         0.18         C         146         0         200           Benzolg, h]iperylene         2.3         C         162         0         200           Acenaphthene         1074.1         N         147         3         3800           Pyrene         537.0         N         163         3         5200           Arsenic         0.21         C         267         126         1860           Beryllium         0.084         C         40         2         60           Marganese         89.5         N         45         45         24200           Mercury         5.4         N         45         45         24200           Mercury         5.4         N         45         45         24200           Mercury         5.4         N         20         21         5910           Zinc         57         N         10         20         2200           Action         7         N         10<		Benzo[k]fluoranthene	0.93	C	131	1	700	131	100.0%
Dibenz(a,h)anthracene         0.045         C         146         0         200           Indeno[1,2,3-cd]pyrene         0.18         C         146         0         200           Benzo[g,h,i]perylene         2.3         C         162         0         200           Acenaphthene         1074.1         N         147         3         3800           Arsenic         0.21         C         267         126         1800           Arsenic         0.21         C         267         126         1850           Beryllium         0.084         C         40         2         60           Marganese         89.5         N         45         45         1850           Mercury         5.4         N         45         45         24200           Mercury         5.4         N         45         45         24200           Mercury         5.4         N         335         321         100000           Cadmium         9.0         N         296         13600           Selenium         89.5         N         170         20         2200           Antimony         172         N         174	<del></del>	Chrysene	0.049	၁	163	0	100	163	100.0%
Indeno[1,2,3-cd]pyrene         0.18         C         146         0         200           Bernzo[g,h.i]perylene         2.3         C         162         0         200           Acenaphthene         1074.1         N         147         3         3800           Pyrene         537.0         N         163         3         5200           Arsenic         0.21         C         267         126         1860           Beryllium         0.084         C         40         2         60           Manganese         89.5         N         45         24200           Mercury         5.4         N         45         24200           Mercury         5.4         N         298         220         23300           Cadmium         9.0         N         297         296         13600           Selenium         89.5         N         207         296         13600           Anttinony         7.2         N         172         7         17290           Barrium         117         47200         20         220         2200           Nickel         89.5         N         136         57		Dibenz[a,h]anthracene	0.045	ى د	146	0	200	146	100.0%
Benzolg, h, ilperylene         2.3         C         162         0         200           Acenaphthene         1074.1         N         147         3         3800           Pyrene         537.0         N         163         3         5200           Arsenic         0.21         C         267         126         1860           Beryllium         0.084         C         40         2         60           Marcury         89.5         N         45         45         24200           Mercury         5.4         N         335         321         100000           Lead         7.7         N         298         220         23300           Zinc         5370.4         N         297         296         136000           Selenium         89.5         N         297         296         13600           Mickel         338.0         N         170         20         2200           Nickel         338.0         N         172         47200           Copper         662.3         N         144         117         47200           Chromium         89.5         N         10         20		Indeno[1,2,3-cd]pyrene	0.18	၁	146	0	200	146	100.0%
Accnaphthene         1074.1         N         147         3         3800           Pyrene         537.0         N         163         3         5200           Arsenic         0.21         C         267         126         1860           Beryllium         0.084         C         40         2         60           Marganese         89.5         N         45         24200           Mercury         5.4         N         45         24200           Lead         7.7         N         298         220         23300           Cadmium         9.0         N         291         236         136000           Selenium         89.5         N         207         91         2500           Antimony         7.2         N         170         20         2200           Nickel         358.0         N         136         57         17290           Barium         1233.1         N         136         57         17290           Copper         662.3         N         136         57         1500           Silver         89.5         N         136         57         500		Benzo[g,h,i]perylene	2.3	၁	162	0	200	142	87.7%
Pyrene         537.0         N         163         3         5200           Arsenic         0.21         C         267         126         1860           Beryllium         0.084         C         40         2         60           Manganese         89.5         N         45         24200         60           Mercury         5.4         N         45         24200         60           Lead         7.7         N         298         220         23300           Lead         7.7         N         298         220         23300           Zinc         Selenium         89.5         N         291         215         5910           Selenium         89.5         N         207         91         2500         200           Antimony         7.2         N         170         20         2200         2200           Nickel         358.0         N         136         57         17290         200           Bartium         1253.1         N         124         117         47200         20           Copper         6600         N         136         39         1540         20		Acenaphthene	1074.1	N	147	3	3800	-	0.7%
Arsenic         0.21         C         267         126         1860           Beryllium         0.084         C         40         2         60           Manganese         89.5         N         45         45         24200           Marcury         5.4         N         335         321         100000           Lead         7.7         N         298         220         23300           Cadmium         9.0         N         291         215         5910           Zinc         5370.4         N         291         215         5910           Selenium         89.5         N         207         91         2500           Antimony         7.2         N         170         20         2200           Nickel         358.0         N         176         20         2200           Nickel         358.0         N         144         117         47200           Copper         662.3         N         136         39         1540           Chromium         89.5         N         100         20         20		Pyrene	537.0	N	163	3	5200	_	0.6%
m         0.084         C         40         2         60           esc         89.5         N         45         24200           /         5.4         N         335         321         100000           m         9.0         N         298         220         23300           m         9.0         N         291         215         \$910           n         5370.4         N         297         296         136000           n         89.5         N         207         91         2500           y         7.2         N         170         20         2200           y         7.2         N         136         57         17290           y         1253.1         N         1344         117         47200           s         662.3         N         136         297         281         66900           m         89.5         N         102         60         60	Trace Metals	Arsenic	0.21	၁	267	126	1860	267	100.0%
ese         89.5         N         45         45         24200           /         5.4         N         335         321         100000           m         7.7         N         298         220         23300           m         9.0         N         291         215         5910           n         5370.4         N         297         296         136000           n         89.5         N         207         91         2500           ny         7.2         N         170         20         2200           n         7.2         N         172         47200           s         7.2         N         144         117         47200           e62.3         N         136         39         1540           m         89.5         N         100         50         50		Beryllium	0.084	၁	40	2	09	40	100.0%
/         5.4         N         335         321         100000           m         7.7         N         298         220         23300           m         9.0         N         291         215         5910           n         89.5         N         297         296         136000           y         7.2         N         207         91         2500           y         7.2         N         170         20         2200           y         7.2         N         170         20         2200           x         7.2         N         136         57         17290           x         662.3         N         144         117         47200           x         662.3         N         136         39         1540           x         107         50         50         50		Manganese	89.5	Z	45	45	24200	45	100.0%
m     7.7     N     298     220     23300       m     9.0     N     291     215     5910       n     5370.4     N     297     296     136000       n     89.5     N     207     91     2500       n     7.2     N     170     20     2200       y     7.2     N     136     57     17290       x     1253.1     N     144     117     47200       x     662.3     N     136     39     1540       x     103     50     50     50		Метсигу	5.4	Z	335	321	100000	334	99.7%
m     9.0     N     291     215     5910       n     5370.4     N     297     296     136000       n     89.5     N     207     91     2500       y     7.2     N     170     20     2200       y     358.0     N     136     57     17290       x     1253.1     N     144     117     47200       x     662.3     N     136     39     1540       x     103     50     50     50		Lead	7.7	N	298	220	23300	287	96.3%
n         89.5         N         297         296         136000           n         89.5         N         207         91         2500           ny         7.2         N         170         20         2200           358.0         N         136         57         17290           1253.1         N         144         117         47200           662.3         N         297         281         66900           M         136         39         1540           m         89.5         N         100         50         50	<del></del>	Cadmium	9.0	Z	291	215	\$910	250	85.9%
n         89.5         N         207         91         2500           yy         7.2         N         170         20         2200           358.0         N         136         57         17290           1253.1         N         144         117         47200           662.3         N         297         281         66900           mm         89.5         N         102         50         50		Zinc	5370.4	Z	297	296	136000	241	81.1%
y         7.2         N         170         20         2200           358.0         N         136         57         17290           1253.1         N         144         117         47200           662.3         N         297         281         66900           103         89.5         N         136         39         1540           103         89.5         N         102         60         60	_	Selenium	89.5	Z	207	91	2500	158	76.3%
358.0 N 136 57 17290 1253.1 N 144 117 47200 662.3 N 297 281 66900 110 89.5 N 136 39 1540 110 6000 110 600 6000 110 600 6000 110 600 60		Antimony	7.2	Z	170	70	2200	114	67.1%
662.3 N 297 281 66900 mm 89.5 N 136 39 1540		Nickel	358.0	Z	136	57	17290	22	61.8%
662.3 N 297 281 66900 mm 89.5 N 136 39 1540 mm		Barium	1253.1	Z	141	117	47200	25	58.3%
ium 89.5 N 136 39 1540		Copper	662.3	Z	297	281	00699	173	58.2%
89 S		Silver	89.5	Z	136	39	1540	99	48.5%
107 69 670		Chromium	89.5	z	102	88	620	4	43.1%

		(Page 5 of 5)	(Page 5 of 5)	CAEENING CO	NCEN INC	2101		
		Screening Tissue Concentration (STC)	STC	Total	Number	Maximum Concentration	Total Number of STC	Frequency of STC
Chemical Group	Chemical	(μg/kg) <sup>a</sup>	Classification <sup>b</sup>	Measurements	Detected	(µg/kg) <sup>a</sup>	Exceedances	Exceedances
Radionuclides	Americium 241	0.00084	၁	33	0	0.0135	33	100.0%
	Cesium 137	0.0072	၁	33	2	90.0	33	100.0%
	Europium 152	960.0	၁	33	0	0.2	33	100.0%
	Europium 154	0.067	၁	33	0	0.125	33	100.0%
	Plutonium 238	0.00092	၁	33	1	0.011	33	100.0%
	Plutonium 239/240	0.00088	၁	33	16	0.0055	31	93.9%
	Cobalt 60	0.013	၁	33	0	0.075	. 91	48.5%
<sup>a</sup> All concentrations are reported in units of μg/k	ported in units of µg/kg wet weight, exc	kg wet weight, except for radionuclides. Radionuclide concentrations are reported as pCi/g wet weight	Radionuclide cor	centrations are r	eported as pC	i/g wet weight.		
d - N - M - M - M								

	TABLE 8. CHEMICALS 1	CHEMICALS THAT DID NOT EXCEED TISSUE SCREENING CONCENTRATIONS	ED TISSUE SCREE	NING CONCEN	FRATIONS		
		Screening Tissue				Maximum	Total Number
Chemical Group	Chemical	Concentration (STC) (ug/kg) <sup>a</sup>	STC Classification <sup>b</sup>	Total Measurements	Number Detected	Concentration (ug/kg) <sup>a</sup>	of STC Exceedances
Pesticides							
Dinitroanilines	Isopropalin	268.5	z	18	0	1.25	0
-	Trifluralin	134.3	Z	18	1	7.16	0
Organochlorines	Dacthal	8950.6	Z	81	6	50	0
	Pentachloronitrobenzene	1.4	C	18	0	1.25	0
Organophosphates	Chlorpyrifos	53.7	N	18	-	3.44	0
	Malathion	358.0	Z	72	2	110	0
	Parathion	107.4	Z	72	3	26	0
Radionuclides	Europium 155	0.45	၁	33	0	0.25	0
Semi-Volatiles	1,2,4,5-Tetrachlorobenzene	5.4	Z	18	2	1.61	0
	1,2-Dichlorobenzene	1611.1	Z	129	0	250	0
	1,3-Dichlorobenzene	1593.2	z	129	0	250	0
	2,4,5-Trichlorophenol	1790.1	Z	56	0	1250	0
	2-Methylphenol	895.1	Z	129	0	326	0
	4-Bromophenyl phenyl ether	1038.3	Z	129	0	250	0
	Benzoic acid	71604.9	Z	56	3	2500	0
	Benzyl Alcohol	17901.2	Z	56	2	250	0
	Di-n-butylphthalate	1790.1	Z	129	10	1550	0
	Diethyl phthalate	14321.0	Z	129	0	250	0
	Dimethyl phthalate	1790.1	Z	129	0	326	0
	Pentachlorobenzene	14.3	Z	18	1	1.25	0
	Phenol	10740.7	z	129	10	2000	0
Semi-Volatile PAHs	2-Chloronaphthalene	1432.1	Z	129	0	250	0
	Anthracene	5370.4	z	163	0	100	0
	Fluoranthene	716.0	z	163	10	100	0
	Fluorene	716.0	Z	163	3	100	0
	Naphthalene	716.0	Z	164	26	200	0
a A II		519.1	Z	163	12	100	0
All concentrations are reported in units of ug/l	cg wet weight,	except for radionuclides.	. Radionuclide concentrations are reported as pCi/g wet weight	ntrations are repor	nted as pCi/g v	wet weight.	

Analyte Group	Analytical Method
Dioxins/Furans	EPA 1613B
Coplanar PCBs	NFCRC C5.181
Pesticides/PCBs	EPA 8081
Semivolatile organics	EPA 8270
PAHs	EPA 8270 with selected ion monitoring (SIM)
Metals	EPA 6010A

A contract laboratory will be responsible for processing the collected fish samples and for analysis of dioxin/furans and coplanar PCBs. The U.S. EPA Manchester Laboratory in Port Orchard, Washington will be responsible for all other analyses.

The resources allocated for chemical analyses do not presently provide for the analysis of radionuclides in tissue. Design Conference attendees recommended that analysis of radionuclides be included in the Phase II study. EPA staff are currently trying to determine whether an EPA laboratory can perform these analyses; if so, they will be included in the study design. If an EPA laboratory cannot provide these analyses, radionuclides will not analyzed. This issue is expected to be resolved prior to the preparation of the draft sampling and QA/QC plan.

#### 3.0 RECOMMENDATIONS

Design Conference attendees provided several recommendations that address a variety of issues relevant to the Phase II study and broader objectives for assessing the impacts of toxic contaminants and habitat degradation on fish stocks, ecological health, and human health. These recommendations are listed below.

#### 3.1 GENERAL COMMENTS

- Studies should be designed with the goal of providing information that will allow better protection of natural resources.
- Ecological impairment should be evaluated. An ecological risk design conference should be held to develop specific objectives and a study plan for assessing ecological impairment.
- Regulatory agencies should coordinate their risk assessment activities to ensure that the public receives a consistent message.
- The methodology for conducting an assessment of human health for the CRITFC member tribes should be clearly delineated, as well as the form in which this information will be conveyed to the public.

## 3.2 COMMENTS SPECIFIC TO THE PHASE II STUDY

- A sampling and QA/QC document should be prepared for the Phase II study that includes a schedule for the project collection activities, report due dates, and peer review.
- Radionuclides should be analyzed.

- Composite samples should consist of fish within a specified size range. It is recommended that the size range include the larger fish within a given population, since these fish may contain higher contaminant burdens.
- A detailed study plan should be developed for determining biota-sediment-accumulation factors (BSAFs) for the Columbia River Basin.
- The sampling and QA/QC plan for the Phase II study should include guidance on selecting alternative species, or locations, if sufficient numbers of the target species can not be collected.
- If resources are insufficient to collect all of the samples included in this study design, it is recommended that the following samples, in order listed, be eliminated: largescale sucker at site 98, fall chinook at site 96.
- Any observed external anomalies in the fish collected should be collected.
- The inclusion in the study design of pathological analyses and measurement of fish hormone levels of fish collected should be considered.

#### 4.0 REFERENCES

CRITFC (Columbia River Inter-Tribal Fish Commission). 1994. A fish consumption survey of the Umatilla, Nez Perce, Yakima, and Warm Springs Tribes of the Columbia River Basin. CRITFC Technical Report No. 94-3. Portland, Oregon.

Hatcher, L. 28 September 1994. Personal communication (Telephone conversation with Dr. Steve Ellis, Tetra Tech, Inc., Redmond, Washington). Yakima Indian Nation, Fisheries Resource Management, Toppenish, Washington.

Tetra Tech, Inc., 1993. Reconnaissance survey of the lower Columbia River. Task 6: Reconnaissance report (3 volumes). Prepared for Columbia River bi-State Committee. Tetra Tech, Inc., Redmond, Washington. Volume I - 454 pp + appendices. Volume 2 - Appendix A, Data Validation Reports - 361 pp. Volume 3 - Data Tables, Appendices B, C, D, and E - 314 pp.

Tetra Tech, Inc., 1994a. Assessing human health risks from chemically contaminated fish in the lower Columbia River. Sampling and Quality Assurance/Quality Control Plan. Prepared for the Columbia River Bi-State Program. Tetra Tech, Inc., Redmond, Washington.

Tetra Tech, Inc., 1994b. Lower Columbia River backwater reconnaissance survey (3 volumes). Prepared for the Columbia River Bi-State Program. Tetra Tech, Inc., Redmond, Washington.

US Environmental Protection Agency, 1993a. Interim report on data and methods for assessment of 2,3,7,8-tetrachlorodibenzo-p-dioxin risks to aquatic life and associated wildlife. EPA/600/R-93/055. US environmental Protection Agency, Office of Research and Development, Washington, DC.

US Environmental Protection Agency, 1993b. Guidance for assessing chemical contaminant data for use in fish advisories. Volume I: Fish sampling and analysis. EPA 823-R-93-002. US Environmental Protection Agency, Office of Water, Washington, DC.

Washington Department of Fisheries/Oregon Department of Fish and Wildlife (WDF/ODFW), 1993. Status Report. Columbia River Fish Runs and Fisheries, 1938-1992. Washington Department of Fisheries, Olympia, Washington. Oregon Department of Fish and Wildlife, Portland, Oregon.

Washington Department of Fisheries/Oregon Department of Fish and Wildlife (WDF/ODFW), 1994. Joint staff report concerning commercial seasons for spring chinook, sturgeon, shad, and other fisheries and miscellaneous regulations for 1994. Joint Columbia River Management Staff. Washington Department of Fisheries, Olympia, Washington. Oregon Department of Fish and Wildlife, Portland, Oregon.